

Microcirculation and oxidative stress

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Accepted by Dr. Helmut Sies

(Received 19 May 2007; in revised form 10 September 2007)

Abstract

The microcirculation is a complex and integrated system, transporting oxygen and nutrients to the cells. The key component of this system is the endothelium, contributing to the local balance between pro and anti-inflammatory mediators, hemostatic balance, as well as vascular permeability and cell proliferation. A constant shear stress maintains vascular endothelium homeostasis while perturbed shear stress leads to changes in secretion of vasodilator and vasoconstrictor agents. Increased oxidative stress is a major pathogenetic mechanism of endothelial dysfunction by decreasing NO bioavailability, promoting inflammation and participating in activation of intracellular signals cascade, so influencing ion channels activation, signal transduction pathways, cytoskeleton remodelling, intercellular communication and ultimately gene expression. Targeting the microvascular inflammation and oxidative stress is a fascinating approach for novel therapies in order to decrease morbidity and mortality of chronic and acute diseases.

Keywords: Microcirculation, oxidative stress, endothelial dysfunction, antioxidants

Abbreviations: ACE-I, angiotensin-converting enzyme inhibitors; ADMA, asymmetric dimethylarginine; AGEs, advanced glycation products; AP-1, Activator protein-1; BH4, Tetrahydrobiopterin; BMC, bone-marrow cells; cNOS, Constitutive NOS; COX, Cyclooxygenase; CRP, C-reactive protein; CVD, Cardiovascular damage; eNOS, endothelial NOS; ERK, Extracellular signal-regulated kinase; ET-1, Endothelin-1; ET-A, Endothelin-A receptor; HPV, Hypoxic pulmonary vasoconstriction; JNK, Jun N-terminal kinase; KATP, Potassium-Adenosine triphosphate sensitive K+ channels; Kv, Voltage-dependent potassium channels; ICAM-1, Intercellular adhesion molecule-1; IL-1, Interleukin-1; IFN-γ, interferongamma; iNOS, Inducible NOS; I/R, Ischemia reperfusion injury; LDL, Low density lipoprotein; LTB4, Leukotriene B4; MAPK, Mitogen-activated protein kinase; MCP-1, Monocyte chemotactic protein-1; MMPs, Matrix metalloproteinases; MPO, Myeloperoxidase; NADPH, Reduced nicotinamide adenine dinucleotide phosphate; NO, Nitric oxide; NOS, Nitric oxide synthase; NF-κB, Nuclear factor-κB; nNOS, neuronal NOS; PAF, Platelet-activating factor; PAI-1, Plasminogen activator inhibitor-1; PGI2, Prostacyclin; PI3 K, phosphatidyl-inositol-3-kinase; ROS, Reactive oxygen species; SOD, superoxide dismutase; $TNF-\alpha$, Tumour necrosis factor alpha; TRP, Transient receptor potential channels; TXA2, Thromboxane; VCAM-1, Vascular cell adhesion molecule-1.

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Introduction

The microcirculation is a complex system delivering oxygen in order to meet the cellular oxygen demand. The key component of this system is the endothelium, mediating, under physiologic conditions, several functions to ensure a normal homeostasis.

A dysfunction of normally protective endothelium may contribute to initiation and progression of several diseases, including cardiovascular damage associated to hypercholesterolemia, hypertension, diabetes, ischemia/reperfusion injury and sepsis [1–6]. Increased oxidative stress, resulting from an exceeding production of reactive oxygen species (ROS) and other oxidants, plays an important role in determining microvascular injury [7].

The goal of this review is to highlight the complex interactions between microcirculation and oxidative stress in some acute and chronic diseases.

Microcirculation

The microcirculation is a hidden organ consisting of the smallest blood vessels, including resistance arterioles, capillaries and venules. It is a complex and integrated system, transporting oxygen and nutrients to the cells. The endothelial cells, forming the inner lining of all blood vessels, represent the main cell types of the microcirculation.

Endothelial physiology

The endothelium, an active biologic organ, contributes to the local balance between pro- and antiinflammatory mediators, hemostatic balance, as well as vascular permeability and cell proliferation [8]. Normal endothelial cells show vasodilator, anti-coagulant and anti-adhesive properties.

The endothelium mediates the vasomotor tone of the microcirculation by release of vasodilators (nitric oxide, prostacyclin, bradykinin and endotheliumderived hyperpolarizing factor) and vasoconstrictors (endothelin-1, angiotensin II and thromboxane) factors. Nitric oxide (NO) plays a predominant role in vasodilation [9]. NO is synthesized from the aminoacid L-arginine in a reaction catalysed by a family of nitric oxide synthases (NOSs), requiring tetrahydrobiopterin (BH4) as cofactor, and leads to relaxation of smooth muscle cells by increasing intracellular cyclic guanosine-monophosphate levels.

The small physiologic amounts of NO generated by the constitutive NOSs (neuronal NOS, nNOS, NOS I and endothelial NOS, eNOS, NOS III) are responsible for most of its beneficial effects (vasodilation, inhibition of platelet aggregation and leukocyte adhesion to the endothelium). A constant production of NO contributes to regulation of arterial systemic pressure by maintaining a continuous vasodilator

tone as shown by hypertensive phenotype in eNOS knockout mouse [10].

The inducible NOS (iNOS, NOS II), activated by inflammatory stimuli, produces larger and more persistent concentration of NO, leading to most of its detrimental actions: hypotension, negative inotropic effect, pro-oxidant properties, apoptosis, mediation of the effects of cytokines, cytotoxic innate immunity [11].

The most important physiologic factor for NO synthesis is shear stress, that is a tangential distortion of the endothelial cells produced by blood flow [12]. NO is also released in response to pharmacological agonists such as acetylcholine.

The counterpart of NO is endothelin-1 (ET-1), which causes vasoconstriction, smooth muscle cell proliferation by activation of endothelin-A receptor (ET-A) and release of inflammatory mediators such as interleukin-1 (IL-1), IL-6 and IL-8.

A constant shear stress maintains vascular endothelium homeostasis, preventing cell apoptosis/prolicoagulation, leukocyte adhesion and atherogenesis [13,14], while perturbed shear stress leads to changes in secretion of vasoactive factors as well as in gene expression by activating different signal transduction pathways [15]. For example, acute loss of shear stress, as observed in ischemia, results in membrane depolarization of lung microvascular endothelial cells, secondary to adenosine triphosphate sensitive K⁺ (K_{ATP}) channels closure; this response is followed by activation of endothelial nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, ROS production, activation of transcriptor factors such as nuclear factor-κB $(NF-\kappa B)$ and activator protein-1 (AP-1) and increase of intracellular Ca²⁺, with enhanced eNOS activity and NO release (Figure 1). Ultimately, the endothelial cells try to restore the blood flow by promoting angiogenesis and vasodilation [15,16]. In pulmonary endothelium, calcium influx, via voltage-gated T-Type calcium channels, promotes secretion of von Willebrand factor and expression of P-selectin, so influencing hemostasis and inflammation [17].

The tone of microcirculation is tightly coupled to parenchymal oxygen consumption, resulting in vasodilation or vasoconstriction according to the actual interstitial Po₂ concentration [18]. Endothelial cells might act as oxygen sensor and, when exposed to hypoxia, mediate vasodilation by increased production of NO, prostacyclin (PGI₂) or by activation of K_{ATP} channels in smooth muscle cells with subsequent hyperpolarization and reduced calcium inflow [19,20]. Conversely, hypoxic pulmonary vasoconstriction (HPV) is a feature of pulmonary circulation in response to alveolar hypoxia in order to preserve gas exchange. ROS may participate in HPV by regulating potassium and/or calcium channels [21,22].



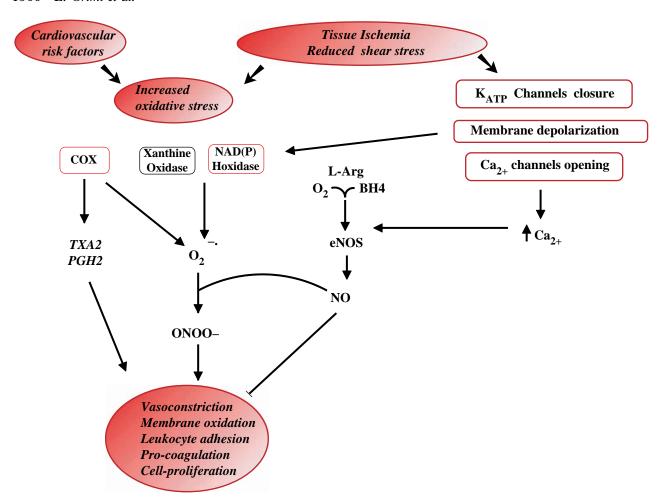


Figure 1. Role of increased oxidative stress and perturbed shear stress on endothelial dysfunction. Main source of ROS are NADPH oxidase, xanthine oxidase and cyclooxygenase (COX). The superoxide anion (O2 - ·) scavenges NO to form peroxynitrite (ONOO). Acute loss of shear stress can result in potassium-adenosine triphosphate sensitive K+ (KATP) channels closure with membrane depolarization, leading on one side to activation of endothelial NADPH oxidase, ROS generation and on the other side to increase of intracellular Ca²⁺ by activation of voltage dependent Ca2+ channels with activation of endothelial NO synthase and NO production in the attempt to restore blood flow.

Endothelial dysfunction

An impairment of endothelial-mediated vasodilation characterizes endothelial dysfunction; it is the result of reduced NO production, mainly due to its inactivation by ROS and increased release of vasoconstrictor factors [23-25]. L-arginine and tetrahydrobiopterin deficiencies cause NOS dysfunction, affecting NO production and increasing the concentration of oxidants such as superoxide and hydrogen peroxide, a phenomenon described as 'NOS uncoupling' [26,27].

Endothelial dysfunction includes a pro-inflammatory, pro-coagulant and proliferative condition, called endothelial activation. Different signals can activate the endothelial cells: cell-to-cell interactions, soluble mediators and hemodynamic forces [28]. Haematic hyperviscosity and reduced erythrocyte deformability with worse hemorheological parameters can increase the risk of ischemia [29].

Dysfunction of the normally protective endothelium is a key component in several diseases, including atherosclerosis, hypertension, diabetes, ischemia reperfusion injury and sepsis [30-33]. Specialized microvasculature is present in the central nervous system where endothelial cells and astrocytes participate in the blood-brain barrier. There is increasing interest in elucidating the relation between the integrins and matrix adhesion receptors and extracellular matrix in the regulation of cerebral vascular permeability as novel therapeutic target in conditions such as cerebral ischemia and multiple sclerosis [34].

The identification of biological markers of endothelial dysfunction could help for an early diagnosis and new effective treatment. Some of the most studied indexes are citrulline/arginine ratio, an index of NOS activity, asymmetric dimethylarginine (ADMA), a potent endogenous inhibitor of NO, synthesis endothelial progenitor cells, myeloperoxidase (MPO) and C-reactive protein (CRP) [35-41].



Oxidative stress

Oxidative stress describes an imbalance in production of free radical species and their effective removal by antioxidants and scavenger enzymes [42]. ROS are normally produced during cellular respiration and inflammatory defense mechanisms [43]. There are several scavenger systems including enzymes such as superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase, a selenium dependent enzyme (GSH-Px), and non-enzymatic antioxidants such as vitamin E, vitamin C, beta-carotene and hemebinding proteins [44].

ROS deplete the cellular levels of NO, increase the expression of adhesion molecules (P-selectin), lipid inflammatory mediators such as platelet-activating factor (PAF), leukotriene B4 (LTB4) and cytokines such as IL-8 [7,25], so contributing to endothelial dysfunction. The superoxide anion (O_2^-) scavenges NO to form peroxynitrite, triggering different proinflammatory signals and inhibiting endothelial repair and preventing angiogenesis [45,46].

Exogenous sources of ROS are phagocytes as well as circulating enzymes such as xanthine oxidase. Endothelial cells can also generate ROS in response to tumour necrosis factor alpha (TNF- α), IL-1 β , platelet-derived growth factor. Endogenous sources of ROS, such as cyclo-oxygenase and NADPH oxidase, might lead to more subtle effects other than cell injury [13]. In fact, ROS might act as second messenger and have a significant effect on the vascular cells signal transduction pathways involving mitogen-activated protein kinase (MAPK), extracellular signal-regulated kinase (ERK), Jun N-terminal kinase (JNK) as well NF- κ B, activator protein-1 activation (AP-1), so influencing gene expression, cytoskeleton organization, cross-talk communication and ion channels activation.

ROS can mediate the endothelial response to mechanical stress, converting the cellular mechanical distortion into biological signals, a process called mechanotransduction [13,15]. NADPH oxidase might represent a specific targeting of signalling modulation and inhibition [47] (Figure 1).

In the pulmonary microcirculation, ROS can induce contraction of pericytes, that surround the endothelial cells, resulting in disruption of the microvasculature barrier and increased permeability [48]. In pulmonary endothelial cells, increased cytosolic calcium level, regulated by T-Type calcium channel and recent identified transient receptor potential channels (TRP) activated by inflammatory mediators, triggers cytoskeleton remodelling responsible for endothelial cell barrier disruption. The inhibition of these channels represents a novel anti-inflammatory strategy [17,49-51].

Ageing

Cell senescence is associated to endothelial dysfunction [52]. Senescent vascular cells show an impairment of endothelium-dependent vasodilation secondary to reduced eNOS activity, NO production and increased ROS production. These cells present a pro-coagulant phenotype, due to increased production of thromboxane A₂ (TXA₂), ET-1 and plasminogen activator inhibitor-1 (PAI-1). Ageing is also associated with higher levels of circulatory inflammatory mediators such as TNF- α , advanced glycation products (AGEs), matrix metalloproteinases (MMPs) and inflammatory cells as mast cells that disrupt the integrity of the microvascular endothelium and, ultimately, reduce blood flow [52-56]. All these changes can exacerbate age-related diseases like hypertension, atherosclerosis and diabetes mellitus.

Targeting senescence mechanisms might represent a new therapeutic strategy. Physical activity and caloric restriction can improve age-related modifications to endothelium and reduce the risk of death from cardiovascular diseases in the elderly [57–59].

Cardiovascualr damage and hypercholesterolemia

Hypercholesterolemia is a major risk factor for coronary atherosclerosis and is associated to endothelial dysfunction [60,61]. Increased oxidative stress can contribute to myocardial vascular dysfunction in hypercholesterolemia [62]. The most important effects of ROS include oxidation of low-density lipoprotein (LDL), arteriolar scavenging of the endothelium-derived NO with reduced vasorelaxation and blood cells-venular endothelial cells interaction.

Oxidative modification of LDL may promote endothelial injury with progression of the fatty lesions [63]. Oxidized LDLs antagonize the endothelial production of NO, reducing the expression of eNOS [64,65], decreasing the uptake of L-arginine and enhancing the level of asymmetric dimethylarginine (ADMA) [66]. Oxidative stress, by activation of NF- κ B, can influence the expression of IL-1, vascular cell adhesion molecule-1 (VCAM-1), ICAM-1, pro-angiogenic factors such as ET-1, MMP-2 and MMP-9, thus contributing to vasa vasorum and atherosclerotic plaque neovascularization [67-69]. Angiogenesis can contribute to progression as well as to vulnerability of atherosclerotic lesions [70]. The vasodilation in response to substances like acetylcholine (Ach) is diminished, as a result of increased ROS and decreased bioavailability of NO in arterioles [71,72].

In post-capillary venules, oxidative stress promotes adhesion molecules expression with platelet and leukocyte recruitment, a response attenuated by NO



supplementation [73,74]. Platelets play a complex role, linking thrombosis, inflammation and immune response [75]. Platelet activation produces peroxynitrite, which increases thromboxane levels and inhibits the mechanisms of endothelial repair. The interaction of platelets and endothelial cells promotes the release of IL-8, monocyte chemotactic protein-1 (MCP-1) and ultimately monocyte activation [35].

MPO, released by neutrophils and monocytes, is a powerful generator of oxidizing, nitrating and chlorinating species with proinflammatory and proatherogenic properties. It can oxidize LDL cholesterol, activate metalloproteinases and decrease NO availability. MPO is involved in destabilization and rupture of the atherosclerotic plaque and associated to adverse outcome in patients with acute coronary syndrome [39,40].

Cytokines, such as interferon- γ (IFN- γ), TNF- α , IL-1, released by activated leukocytes, increase oxidative stress in venules [76,77] and arteriolar dysfunction in hypercholesterolemia [78].

Endothelial dysfunction induced by oxidized LDL can be reversed by L-arginine [79,80], pomegranate juice containing polyphenolic antioxidants [81,82], flavanol-rich cocoa [83,84], antioxidants like vitamins C and E [80,85,86], inhibition of the renin angiotensin system [87] and cholesterol lowering statins [88-90]. Metabolic treatment with antioxidant and L-arginine associated to moderate physical exercise in experimental models of hypercholesterolemia showed protective effects, with reduction of atherosclerotic lesion formation, plaque rupture and prolonged survival [91,92]. Interestingly, chronic exercise elicited an increase in production of NOS expression and improved scavenger activities [93].

Notably, the beneficial effects of chronic antioxidant supplementation in hypercholesterolemia are not confirmed under normal conditions, where it can impair myocardial perfusion and coronary endothelial function by an increased level of oxidative stress in the arterial wall [94]. Therefore, high-dose antioxidant vitamins in healthy subjects with presumed low oxidative stress might be detrimental.

A new promising therapeutic approach consists of targeting the processes involved in angiogenesis. Statins decrease vascular wall oxidative stress by eNOS activation with increased NO bioavailability and a direct antioxidant activity [88,89]. Moreover, they reduce angiogenesis and vasa vasorum neovascularization [90].

The blockade of the endothelin system, which is upregulated in hypercholesterolemia, showed protective effects on renal microvasculature [95].

Over the past few years, CRP has been investigated not only as an inflammatory marker but also as a possible mediator of atherosclerosis [96]. In particular, CRP can influence the microcirculation by eNOS downregulation [97], increasing ET-1 [98]

and upregulating angiotensin type 1 receptor in smooth muscle cells [99]. CRP also promotes ROS production and upregulation of chemokines and endothelial cell adhesion molecules, so contributing to a proinflammatory and proatherosclerotic environment [100,101]. CRP could represent not only a diagnostic marker during primary prevention of cardiovascular diseases [102] but also a new therapeutic target in cardiovascular protection [103,104].

Cardiovascular damage and hypertension

Hypertension is a significant cardiovascular risk factor, associated to endothelial dysfunction and oxidative stress. An increased production of ROS and a reduced level of SOD are reported in both animal [105-109] and human studies [110]. Oxidative stress participates in increasing systemic arterial pressure, reducing NO availability and vasodilation [111,112]. NADPH oxidase and xanthine oxidase are the main sources of ROS in hypertension [113,114]. Oxidative stress is involved in remodelling of myocardial microvascular architecture and subsequent development of left ventricular hypertrophy [115,116]. Hypertension, combined to hypercholesterolemia, accentuates oxidative stress, showing a synergistic deleterious effect on myocardial microvascular dysfunction [117,118].

Antioxidant supplementation with high dose of vitamins C and E can reduce endothelial dysfunction and improve myocardial perfusion in early hypertension [119]. Simvastatin showed a protective effect, preventing myocardial microvascular remodelling and hypertrophy in experimental renovascular hypertension [120].

Cardiovascular damage and diabetes

Diabetes mellitus is a major cardiovascular risk factor, associated with increased morbidity mortality [121]. Endothelial dysfunction with impaired endothelium-dependent vasodilation has been documented in type 1 [122] and type 2 diabetes [123] as well as in the syndrome of insulin resistance [124]; it also plays an important role in the development of macro and microvascular disease [125]. Suggested cellular mechanisms of impaired endothelium-dependent vasodilation include decreased production of vasodilators (NO, prostacyclin, endothelium-derived hyperpolarizing factor) and increased release of vasoconstrictors (thromboxane, ET-1) [4]. Disturbance in voltage-gated K⁺ channel function [126] and impaired vasodilation to hypoxia can contribute to microvascular dysfunction [127]. A defect of insulin action on phosphatidyl-inositol-3-kinase (PI3-K) pathway that normally mediates the vasodilator effect of insulin by increasing eNOS



gene expression and NO bioavailability can explain the vascular dysfunction in insulin resistance [128, 129].

Hyperglycemia-induced oxidative stress plays a central role in endothelial dysfunction [130,131]. Hyperglycemia, the hallmark of diabetes, increases superoxide production by NADPH depletion with glutathione regeneration impairment and activation of vascular NADPH oxidase due to increased AGE production and protein kinase C activation [131]. Other possible sources of ROS include uncoupled eNOS, xanthine oxidase and mitochondria [4]. Hyperglycemia and oxidative stress, through increased LDL oxidation, can accelerate the atherosclerotic disease in diabetic patients [132].

Antioxidants (N-acetylcysteine, vitamins E and C) can restore endothelial function [133-135]. In an experimental model of metabolic syndrome, characterized by insulin resistance, pomegranate fruit extract, rich in polyphenolic antioxidants, reduced the expression of oxidation-sensitive genes at the sites of perturbed shear-stress and increased eNOS expression [136]. However, antioxidant therapy does not reduce cardiovascular complications in diabetic patients [137].

ACE inhibitors can decrease NADH oxidase activity and free radicals by inhibition of angiotensin II [125]. Statins improved endothelial dysfunction in diabetes, independently of changes in cholesterol levels [138,139].

Cardiovascular damage and ischemia/ repurfusion injury

Ischemia/reperfusion injury (I/R) defines the cellular structural changes occurring after reperfusion of ischemic tissue [5]. Endothelial cells exposed to the deleterious effects of ischemia and reperfusion show depletion of energy stores, altered ion distribution, membrane depolarization, increased hypoxanthine level, increased membrane fluidity, cellular swelling and cytoskeleton derangement [140].

The components of the microcirculation including arterioles, capillaries and venules manifest different responses to I/R injury.

I/R results in severe dysfunction of the endothelium in arterioles characterized by impaired endotheliumdependent vasodilation and increased arterial resistance. A reduction in NO release is responsible for impaired arteriolar vasodilation [141-144]. The mechanisms responsible for this inhibition are still unclear. Increased activity of arginase [145], enzyme competing with NOS for the substrate L-arginine, and depletion of BH4, cofactor of NOS [146], have been described following I/R injury. A burst of oxidant production at the onset of the reperfusion can impair the NO-dependent vasodilation by direct

inhibition [147] and stimulating leukocyte adhesion molecule, promoting leukocytes activation, chemotaxis and ultimately leading to further ROS production. Interestingly, inhibition of arginase [145], administration of BH4 [148] as well as gene deficiency of leukocyte adhesion molecules (CD11/18, P selectin, ICAM-1) [149] can restore and/or preserve endothelium-dependent NO-mediated relaxation.

The microvascular dysfunction in capillaries includes increased fluid filtration with interstitial edema and less perfused capillaries. Increased permeability of the endothelium rather than elevated intracapillary pressure is the main mechanism for increased filtration [150]; it can be due to NO inhibition [151] as well as to intercellular adhesion disruption secondary to ROS production and energy depletion. Decreased endothelium-dependent vasorelaxation, interstitial fluid accumulation, obstruction and/or narrowing of the vessels by platelet-leukocyte aggregation and leukocyte-endothelial cell adhesion can lead to mechanical blood flow obstruction and capillary malperfusion following the reperfusion of an ischemic organ, known as 'no reflow phenomenon' [152]; it may be clinically evident as persistence and/or worsening of organ dysfunction after the reperfusion.

In the post-capillary venules, endothelium dysfunction is characterized by an intense inflammatory response. ROS play a pivotal role in this response. On reperfusion of ischemic tissues, ROS production is enhanced by high levels of xanthine oxidase converting the intracellular excess of hypoxanthine as a result of ATP degradation [5]. ROS promote activation of circulating leukocytes, platelets, with generation of inflammatory mediators (PAF, LTB, IL-8), cytokine and adhesion molecule expression, resulting in neutrophil mediated tissue injury [153,154]. Potential strategies to prevent I/R, including ischemic pre-conditioning, antioxidant, antileukocyte and anticomplement therapy, did not provide a definite and unequivocal benefit in the clinical settings [140].

Cardiovascular damage and sepsis

Microvascular dysfunction participates in the pathogenesis of sepsis. Pathogens, lipopolysaccharide as well as inflammatory mediators can be responsible for endothelial cell activation and dysfunction [155].

An increased heterogeneity in microvascular blood flow has been described in various experimental models of sepsis [156-159] and in patients with septic shock, using polarized light microscopy [160]. The early phases of sepsis are characterized by reduced number of perfused capillaries, misdistribution of oxygen delivery and subsequent tissue hypoxia [161].



Oxidative stress is the result of activation of phagocytes, production of NO and ROS, release of iron, copper ions, metalloproteins and contributes to experimental microvascular dysfunction in sepsis. ROS can directly disrupt endothelial cells and impair cellular interaction, so promoting microvascular thrombosis and organ dysfunction [162,163].

NO is an important factor for the integrity of microvascular endothelium and blood flow [164]. Bacterial endotoxin or inflammatory cytokines increase iNOS activity. The enhanced production of NO contributes to vascular collapse and myocardial dysfunction, mediating the depressant effects of proinflammatory cytokines (TNF- α , IL-1 β) [165– 168]. Endotoxin produced less hypotension in iNOS deficient mice [169] and in mice treated with a selective pharmacological inhibitor of iNOS [170]. The heterogeneous expression of iNOS in different vascular beds can result in flow shunting [171]. NO could have beneficial effects in sepsis related to counteraction of the released vasoconstrictors substances, to inhibition of leukocyte rolling and adhesion and to inhibition of NF- κ B [172]. Interestingly, myocardial over-expression of NOSIII showed a protective effect against endotoxin-induced myocardial dysfunction [173].

Conclusions

The endothelium represents a dynamic interface with activation of different and highly integrated cellular pathways in response to mechanical and metabolic stimuli. An imbalance between ROS production and NO availability leads to main changes in endothelial function and ultimately to impaired microcirculatory perfusion. Oxidative stress might play a more sophisticated role in endothelial dysfunction other than cellular damage. It can act as an important mediator in the inflammatory cascade and participate in mechanotransduction, transforming the mechanical forces changes determined by perturbed shear stress in biological signals.

Microcirculatory dysfunction is emerging as an important factor in the pathogenesis of acute and chronic diseases. Interestingly, senescence is also associated to increased oxidative stress and endothelial dysfunction, contributing to the exacerbation of age-related cardiovascular disease.

Targeting the oxidant response by antioxidants and modulating specific enzymes (e.g. NO synthases, NADPH oxidase) represents a potential therapeutic strategy.

Recent evidence indicates that treatment with antioxidants and L-arginine combined to bone-marrow cells (BMC) transplantation can provide beneficial effects beyond those achieved by BMC transplantation alone in the healing process of the

injured cardiovascular system [174]. There is growing interest on the protective effects of statins on the endothelial dysfunction, beyond their lipid lowering action. Healthy lifestyle, including regular physical activity and balanced diet, is an easy practice to reduce the endothelium age-related modifications.

References

- [1] Davignon J, Ganz P. Role of endothelial dysfunction in atherosclerosis. Circulation 2004;109:III27-III32.
- [2] Suematsu M, Suzuki H, DeLano FA, Schmid-Schonbein GW. The inflammatory aspect of the microcirculation in hypertension: oxidative stress, leukocytes/endothelial interaction, apoptosis. Microcirculation 2002;9;259-276.
- [3] DeLano FA, Balete R, Schmid-Schonbein GW. Control of oxidative stress in microcirculation of spontaneously hypertensive rats. Am J Physiol Heart Circ Physiol 2005;288: H805-H812.
- [4] Rask-Madsen C, King GL. Mechanisms of disease: endothelial dysfunction in insulin resistance and diabetes. Nat Clin Pract Endocrinol Metab 2007;3:46–56.
- [5] Carden DL, Granger DN. Pathophysiology of ischaemiareperfusion injury. Pathophysiology of ischaemia-reperfusion injury. J Pathol 2000;190:255-266.
- [6] Bateman RM, Sharpe MD, Ellis CG. Bench-to-bedside review: microvascular dysfunction in sepsis--hemodynamics, oxygen transport, and nitric oxide. Crit Care 2003;7: 359-373.
- [7] Cooper D, Stokes KY, Tailor A, Granger DN. Oxidative stress promotes blood cell-endothelial cell interactions in the microcirculation. Cardiovasc Toxicol 2002;2:165-180.
- [8] Tritto I, Ambrosio G. Spotlight on microcirculation: an update. Cardiovasc Res 1999;42:600-606.
- Moncada S, Higgs EA. The discovery of nitric oxide and its role in vascular biology. Br J Pharmacol 2006;147(Suppl 1):S193-S201.
- [10] Huang PL. Lessons learned from nitric oxide synthase knockout animals. Semin Perinatol 2000;24:87-90.
- [11] Ignarro LJ, Cirino G, Casini A, Napoli C. Nitric oxide as a signaling molecule in the vascular system: an overview. J Cardiovasc Pharmacol 1999;34:879-886.
- [12] Fisher AB, Chien S, Barakat AI, Nerem RM. Endothelial cellular response to altered shear stress. Am J Physiol Lung Cell Mol Physiol 2001;281:L529-L533.
- [13] Ali MH, Schumacker PT. Endothelial responses to mechanical stress: where is the mechanosensor? Crit Care Med 2002;30:S198-S206.
- [14] Resnick N, Yahav H, Schubert S, Wolfovitz E, Shay A. Signalling pathways in vascular endothelium activated by shear stress: relevance to atherosclerosis. Curr Opin Lipidol 2000;11:167-177.
- [15] Fisher AB, Al-Mehdi AB, Manevich Y. Shear stress and endothelial cell activation. Crit Care Med 2002;30: S192-S197.
- [16] Chatterjee S, Levitan I, Wei Z, Fisher AB. KATP channels are an important component of the shear-sensing mechanism in the pulmonary microvasculature. Microcirculation 2006;13:633-644.
- [17] Zhou C, Wu S. T-type calcium channels in pulmonary endothelium. Microcirculation 2006;13:645-656.
- [18] Granger HJ, Goodman AH, Cook BH. Metabolic models of microcirculatory regulation. Fed Proc 1975;34:2025-2030.
- [19] Vallet B. Endothelial cell dysfunction and abnormal tissue perfusion. Crit Care Med 2002;30:S229-S234.



- [20] Daut J, Maier-Rudolph W, von Beckerath N, Mehrke G, Gunther K, Goedel-Meinen L. Hypoxic dilation of coronary arteries is mediated by ATP-sensitive potassium channels. Science 1990;247:1341-1344.
- [21] Moudgil R, Michelakis ED, Archer SL. The role of k+ channels in determining pulmonary vascular tone, oxygen sensing, cell proliferation, and apoptosis: implications in hypoxic pulmonary vasoconstriction and pulmonary arterial hypertension. Microcirculation 2006;13:615-632.
- [22] Waypa GB, Schumacker PT. Hypoxic pulmonary vasoconstriction: redox events in oxygen sensing. J Appl Physiol 2005;98:404-414.
- [23] Napoli C, Ignarro LJ. Nitric oxide and atherosclerosis. Nitric Oxide 2001;5:88-97.
- [24] Napoli C, de Nigris F, Williams-Ignarro S, Pignalosa O, Sica V, Ignarro LJ. Nitric oxide and atherosclerosis: an update. Nitric Oxide 2006;15:265-279.
- [25] Napoli C, de Nigris F, Palinski W. Multiple role of reactive oxygen species in the arterial wall. J Cell Biochem 2001;82:674-682.
- [26] Verma S, Maitland A, Weisel RD, Fedak PW, Pomroy NC, Li SH, Mickle DA, Li RK, Rao V. Novel cardioprotective effects of tetrahydrobiopterin after anoxia and reoxygenation: identifying cellular targets for pharmacologic manipulation. J Thorac Cardiovasc Surg 2002;123:1074-1083.
- [27] Cai H, Harrison DG. Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. Circ Res 2000;87:840-844.
- [28] Collins DM, McCullough WT, Ellsworth ML. Conducted vascular responses: communication across the capillary bed. Microvasc Res 1998;56:43-53.
- [29] Turchetti V, Boschi L, Donati G, Trabalzini L, Forconi S. Impact of hemorheological and endothelial factors on microcirculation. Adv Exp Med Biol 2006;578:107-112.
- [30] Bonetti PO, Lerman LO, Lerman A. Endothelial dysfunction: a marker of atherosclerotic risk. Arterioscler Thromb Vasc Biol 2003;23:168-175.
- [31] Savoia C, Schiffrin EL. Vascular inflammation in hypertension and diabetes: molecular mechanisms and therapeutic interventions. Clin Sci (Lond) 2007;112:375-384.
- [32] Ellis CG, Jagger J, Sharpe M. The microcirculation as a functional system. Crit Care 2005;9:S3-S8.
- [33] Ince C. The microcirculation is the motor of sepsis. Crit Care 2005;9:S13-S19.
- del Zoppo GJ, Milner R. Integrin-matrix interactions in the cerebral microvasculature. Arterioscler Thromb Vasc Biol 2006;26:1966-1975.
- Anwaruddin S, Askari AT, Topol EJ. Redefining risk in acute coronary syndromes using molecular medicine. J Am Coll Cardiol 2007;49:279-289.
- [36] Howard-Alpe GM, Sear JW, Foex P. Methods of detecting atherosclerosis in non-cardiac surgical patients; the role of biochemical markers. Br J Anaesth 2006;97:758-769.
- [37] Ridker PM, Cannon CP, Morrow D, Rifai N, Rose LM, McCabe CH, Pfeffer MA, Braunwald E, Pravastatin or atorvastatin evaluation and infection therapy-thrombolysis in myocardial infarction 22 (PROVE IT-TIMI 22) investigators. C-reactive protein levels and outcomes after statin therapy. N Engl J Med 2005;352:20-28.
- [38] Werner N, Kosiol S, Schiegl T, Ahlers P, Walenta K, Link A, Bohm M, Nickenig G. Circulating endothelial progenitor cells and cardiovascular outcomes. N Engl J Med 2005;353:999-1007.
- [39] Baldus S, Heeschen C, Meinertz T, Zeiher AM, Eiserich JP, Munzel T, Simoons ML, Hamm CW, CAPTURE investigators. Myeloperoxidase serum levels predict risk in patients with acute coronary syndromes. Circulation 2003;108:1440-1445.

- [40] Brennan ML, Penn MS, Van Lente F, Nambi V, Shishehbor MH, Aviles RJ, Goormastic M, Pepoy ML, McErlean ES, Topol EJ, Nissen SE, Hazen SL. Prognostic value of myeloperoxidase in patients with chest pain. N Engl J Med 2003:349:1595-1604.
- [41] Reinhart K, Bayer O, Brunkhorst F, Meisner M. Markers of endothelial damage in organ dysfunction and sepsis. Crit Care Med 2002;30:S302-S312.
- [42] Sies H. Oxidative stress. Introductory remarks. In: Sies H, editor. Oxidative stress. London: Academic Press; 1985. p 1-8.
- [43] Sies H. Biochemistry of oxidative stress. Angew Chem Int Ed 1986;25:1058-1071.
- Sies H. What is oxidative stress? In: Keaney JF, editor. Oxidative stress and vascular diseases, Boston, MA: Kluwer Academic; 2000. p 1-8.
- [45] Crimi E, Sica V, Williams-Ignarro S, Zhang H, Slutsky AS, Ignarro LJ, Napoli C. The role of oxidative stress in adult critical care. Free Radic Biol Med 2006;40:398-406.
- [46] Urbich C, Dernbach E, Aicher A, Zeiher AM, Dimmeler S. CD40 ligand inhibits endothelial cell migration by increasing production of endothelial reactive oxygen species. Circulation 2002;106:981-986.
- [47] Terada LS. Oxidative stress and endothelial activation. Crit Care Med 2002;30:S186-S191.
- [48] Edelman DA, Jiang Y, Tyburski J, Wilson RF, Steffes C. Pericytes and their role in microvasculature homeostasis. J Surg Res 2006;135:305-311.
- [49] Tiruppathi C, Ahmmed GU, Vogel SM, Malik AB. Ca2+ signaling, TRP channels, and endothelial permeability. Microcirculation 2006;13:693-708.
- [50] Cioffi DL, Stevens T. Regulation of endothelial cell barrier function by store-operated calcium entry. Microcirculation 2006;13:709-723.
- [51] Townsley MI, King JA, Alvarez DF. Ca2+ channels and pulmonary endothelial permeability: insights from study of intact lung and chronic pulmonary hypertension. Microcirculation 2006;13:725-739.
- [52] Minamino T, Komuro I. Vascular cell senescence: contribution to atherosclerosis. Circ Res 2007;100:15-26.
- [53] Gerhard M, Roddy MA, Creager SJ, Creager MA. Aging progressively impairs endothelium-dependent vasodilation in forearm resistance vessels of humans. Hypertension 1996;27:849-853.
- [54] Payne GW. Effect of inflammation on the aging microcirculation: impact on skeletal muscle blood flow control. Microcirculation 2006;13:343-352.
- [55] Bearden SE. Effect of aging on the structure and function of skeletal muscle microvascular networks. Microcirculation 2006;13:279-288.
- [56] Poole D, Behnke B, Musch T. Capillary hemodynamics and oxygen pressures in the aging microcirculation. Microcirculation 2006;13:289-299.
- [57] Wang JS, Lan C, Chen SY, Wong MK. Tai Chi Chuan training is associated with enhanced endothelium-dependent dilation in skin vasculature of healthy older men. J Am Geriatr Soc 2002;50:1024-1030.
- [58] Abete P, Cacciatore F, Ferrara N, Calabrese C, de Santis D, Testa G, Galizia G, Del Vecchio S, Leosco D, Condorelli M, Napoli C, Rengo F. Body mass index and preinfarction angina in elderly patients with acute myocardial infarction. Am J Clin Nutr 2003;78:796-801.
- [59] Abete P, Della Morte D, Mazzella F, D'Ambrosio D, Galizia G, Testa G, Gargiulo G, Cacciatore F, Rengo F. Lifestyle and prevention of cardiovascular disease in the elderly: an Italian perspective. Am J Geriatr Cardiol 2006;15:28-34.
- [60] Faxon DP, Fuster V, Libby P, Beckman JA, Hiatt WR, Thompson RW, Topper JN, Annex BH, Rundback JH, Fabunmi RP, Robertson RM, Loscalzo J, American heart



- association. Atherosclerotic vascular disease conference: Writing group III: pathophysiology. Circulation 2004;109:
- [61] Stokes KY, Granger DN. The microcirculation: a motor for the systemic inflammatory response and large vessel disease induced by hypercholesterolaemia? J Physiol 2005;562:
- [62] Rodriguez-Porcel M, Lerman A, Best PJ, Krier JD, Napoli C, Lerman LO. Hypercholesterolemia impairs myocardial perfusion and permeability: role of oxidative stress and endogenous scavenging activity. J Am Coll Cardiol 2001;37:608-615.
- [63] Tsimikas S, Brilakis ES, Miller ER, McConnell JP, Lennon RJ, Kornman KS, Witztum JL, Berger PB. Oxidized phospholipids, Lp(a) lipoprotein, and coronary artery disease. N Engl J Med 2005;353:9-11.
- [64] Steffen Y, Schewe T, Sies H. Epicatechin protects endothelial cells against oxidized LDL and maintains NO synthase. Biochem Biophys Res Commun 2005;331:1277-1283.
- [65] Steffen Y, Jung T, Klotz LO, Schewe T, Grune T, Sies H. Protein modification elicited by oxidized low-density lipoprotein (LDL) in endothelial cells: protection by (-)-epicatechin. Free Radic Biol Med 2007;42:955-970.
- [66] Cooke JP. Asymmetrical dimethylarginine: the Uber marker? Circulation 2004;109:1813-1818.
- [67] Wilson SH, Caplice NM, Simari RD, Holmes DR Jr, Carlson PJ, Lerman A. Activated nuclear factor-kappaB is present in the coronary vasculature in experimental hypercholesterolemia. Atherosclerosis 2000;148:23-30.
- [68] Hajra L, Evans AI, Chen M, Hyduk SJ, Collins T, Cybulsky MI. The NF-kappa B signal transduction pathway in aortic endothelial cells is primed for activation in regions predisposed to atherosclerotic lesion formation. Proc Natl Acad Sci USA 2000;97:9052-9057.
- [69] Herrmann J, Best PJ, Ritman EL, Holmes DR, Lerman LO, Lerman A. Chronic endothelin receptor antagonism prevents coronary vasa vasorum neovascularization in experimental hypercholesterolemia. J Am Coll Cardiol 2002;39:1555-1561.
- [70] Herrmann J, Lerman LO, Mukhopadhyay D, Napoli C, Lerman A. Angiogenesis in atherogenesis. Arterioscler Thromb Vasc Biol 2006;26:1948-1957.
- [71] Napoli C, Lerman LO. Involvement of oxidation-sensitive mechanisms in the cardiovascular effects of hypercholesterolemia. Mayo Clin Proc 2001;76:619-631.
- [72] de Nigris F, Lerman A, Ignarro LJ, Williams-Ignarro S, Sica V, Baker AH, Lerman LO, Geng YJ, Napoli C. Oxidationsensitive mechanisms, vascular apoptosis and atherosclerosis. Trends Mol Med 2003;9:351-359.
- [73] Tailor A, Granger DN. Hypercholesterolemia promotes P-selectin-dependent platelet-endothelial cell adhesion in postcapillary venules. Arterioscler Thromb Vasc Biol 2003;23:675-680.
- [74] Tailor A, Granger DN. Hypercholesterolemia promotes leukocyte-dependent platelet adhesion in murine postcapillary venules. Microcirculation 2004;11:597-603.
- [75] von Hundelshausen P, Weber C. Platelets as immune cells: bridging inflammation and cardiovascular disease. Circ Res 2007;100:27-40.
- [76] Stokes KY, Clanton EC, Bowles KS, Fuseler JW, Chervenak D, Chervenak R, Jennings SR, Granger DN. The role of Tlymphocytes in hypercholesterolemia-induced leukocyte-endothelial interactions. Microcirculation 2002;9:407-417.
- [77] Stokes KY, Clanton EC, Clements KP, Granger DN. Role of interferon-gamma in hypercholesterolemia-induced leukocyte-endothelial cell adhesion. Circulation 2003;107: 2140-2145.
- [78] De Kimpe SJ, Tielemans W, Van Heuven-Nolsen D, Nijkamp FP. Reversal of bradykinin-induced relaxation to

- contraction after interferon-gamma in bovine isolated mesenteric arteries. Eur J Pharmacol 1994;261:111-120.
- [79] Quyyumi AA, Dakak N, Diodati JG, Gilligan DM, Panza JA, Cannon RO 3rd. Effect of L-arginine on human coronary endothelium-dependent and physiologic vasodilation. J Am Coll Cardiol 1997;30:1220-1227.
- [80] de Nigris F, Lerman LO, Ignarro SW, Sica G, Lerman A, Palinski W, Ignarro LJ, Napoli C. Beneficial effects of antioxidants and L-arginine on oxidation-sensitive gene expression and endothelial NO synthase activity at sites of disturbed shear stress. Proc Natl Acad Sci USA 2003;100:1420-1425.
- [81] de Nigris F, Williams-Ignarro S, Lerman LO, Crimi E, Botti C, Mansueto G, D'Armiento FP, De Rosa G, Sica V, Ignarro LJ, Napoli C. Beneficial effects of pomegranate juice on oxidation-sensitive genes and endothelial nitric oxide synthase activity at sites of perturbed shear stress. Proc Natl Acad Sci USA 2005;102:4896-4901.
- [82] de Nigris F, Williams-Ignarro S, Sica V, Lerman LO, D'Armiento FP, Byrns RE, Casamassimi A, Carpentiero D, Schiano C, Sumi D, Fiorito C, Ignarro LJ, Napoli C. Effects of a pomegranate fruit extract rich in punicalagin on oxidation-sensitive genes and eNOS activity at sites of perturbed shear stress and atherogenesis. Cardiovasc Res 2007;73:414-423.
- [83] Heiss C, Dejam A, Kleinbongard P, Schewe T, Sies H, Kelm M. Vascular effects of cocoa rich in flavan-3-ols. JAMA 2003;290:1030-1031.
- [84] Schroeter H, Heiss C, Balzer J, Kleinbongard P, Keen CL, Hollenberg NK, Sies H, Kwik-Uribe C, Schmitz HH, Kelm M. (-)-Epicatechin mediates beneficial effects of flavanolrich cocoa on vascular function in humans. Proc Natl Acad Sci USA 2006;103:1024-1029.
- [85] Rodriguez-Porcel M, Lerman LO, Holmes DR Jr, Richardson D, Napoli C, Lerman A. Chronic antioxidant supplementation attenuates nuclear factor-kappa B activation and preserves endothelial function in hypercholesterolemic pigs. Cardiovasc Res 2002;53:1010-1018.
- [86] Levine GN, Frei B, Koulouris SN, Gerhard MD, Keaney JF Jr, Vita JA. Ascorbic acid reverses endothelial vasomotor dysfunction in patients with coronary artery disease. Circulation 1996;93:1107-1113.
- [87] Anderson TJ, Elstein E, Haber H, Charbonneau F. Comparative study of ACE-inhibition, angiotensin II antagonism, and calcium channel blockade on flow-mediated vasodilation in patients with coronary disease (BANFF study). J Am Coll Cardiol 2000;35:60-66.
- [88] Wassmann S, Laufs U, Muller K, Konkol C, Ahlbory K, Baumer AT, Linz W, Bohm M, Nickenig G. Cellular antioxidant effects of atorvastatin in vitro and in vivo. Arterioscler Thromb Vasc Biol 2002;22:300-305.
- [89] Wilson SH, Simari RD, Best PJ, Peterson TE, Lerman LO, Aviram M, Nath KA, Holmes DR Jr, Lerman A. Simvastatin preserves coronary endothelial function in hypercholesterolemia in the absence of lipid lowering. Arterioscler Thromb Vasc Biol 2001;21:122-128.
- [90] Wilson SH, Herrmann J, Lerman LO, Holmes DR Jr, Napoli C, Ritman EL, Lerman A. Simvastatin preserves the structure of coronary adventitial vasa vasorum in experimental hypercholesterolemia independent of lipid lowering. Circulation 2002;105:415-418
- [91] Napoli C, Williams-Ignarro S, De Nigris F, Lerman LO, Rossi L, Guarino C, Mansueto G, Di Tuoro F, Pignalosa O, De Rosa G, Sica V, Ignarro LJ. Long-term combined beneficial effects of physical training and metabolic treatment on atherosclerosis in hypercholesterolemic mice. Proc Natl Acad Sci USA 2004;101:8797-8802.
- [92] Napoli C, Williams-Ignarro S, de Nigris F, Lerman LO, D'Armiento FP, Crimi E, Byrns RE, Casamassimi A, Lanza



- A, Gombos F, Sica V, Ignarro LJ. Physical training and metabolic supplementation reduce spontaneous atherosclerotic plaque rupture and prolong survival in hypercholesterolemic mice. Proc Natl Acad Sci USA 2006;103: 10479-10484.
- [93] Ignarro LJ, Balestrieri ML, Napoli C. Nutrition, physical activity, and cardiovascular disease: an update. Cardiovasc Res 2007;73:326-340.
- [94] Versari D, Daghini E, Rodriguez-Porcel M, Sattler K, Galili O, Pilarczyk K, Napoli C, Lerman LO, Lerman A. Chronic antioxidant supplementation impairs coronary endothelial function and myocardial perfusion in normal pigs. Hypertension 2006;47:475-481.
- Chade AR, Krier JD, Textor SC, Lerman A, Lerman LO. Endothelin-a receptor blockade improves renal microvascular architecture and function in experimental hypercholesterolemia. J Am Soc Nephrol 2006;17:3394-3403.
- [96] Fruchart JC, Nierman MC, Stroes ES, Kastelein JJ, Duriez P. New risk factors for atherosclerosis and patient risk assessment. Circulation 2004;109:III15-III19.
- [97] Venugopal SK, Devaraj S, Yuhanna I, Shaul P, Jialal I. Demonstration that C-reactive protein decreases eNOS expression and bioactivity in human aortic endothelial cells. Circulation 2002;106:1439-1441.
- [98] Verma S, Li SH, Badiwala MV, Weisel RD, Fedak PW, Li RK, Dhillon B, Mickle DA. Endothelin antagonism and interleukin-6 inhibition attenuate the proatherogenic effects of C-reactive protein. Circulation 2002;105:1890-1896.
- [99] Wang CH, Li SH, Weisel RD, Fedak PW, Dumont AS, Szmitko P, Li RK, Mickle DA, Verma S. C-reactive protein upregulates angiotensin type 1 receptors in vascular smooth muscle. Circulation 2003;107:1783-1790.
- [100] Pasceri V, Willerson JT, Yeh ET. Direct proinflammatory effect of C-reactive protein on human endothelial cells. Circulation 2000;102:2165-2168.
- [101] Verma S, Badiwala MV, Weisel RD, Li SH, Wang CH, Fedak PW, Li RK, Mickle DA. C-reactive protein activates the nuclear factor-kappaB signal transduction pathway in saphenous vein endothelial cells: implications for atherosclerosis and restenosis. J Thorac Cardiovasc Surg 2003;126:1886-1891.
- [102] Napoli C, Lerman LO, de Nigris F, Gossl M, Balestrieri ML, Lerman A. Rethinking primary prevention of atherosclerosis-related diseases. Circulation 2006;114:2517-2527.
- [103] Pepys MB, Hirschfield GM, Tennent GA, Gallimore JR, Kahan MC, Bellotti V, Hawkins PN, Myers RM, Smith MD, Polara A, Cobb AJ, Ley SV, Aquilina JA, Robinson CV, Sharif I, Gray GA, Sabin CA, Jenvey MC, Kolstoe SE, Thompson D, Wood SP. Targeting C-reactive protein for the treatment of cardiovascular disease. Nature 2006;440: 1217-1221.
- [104] Ridker PM. High-sensitivity C-reactive protein, inflammation, and cardiovascular risk: from concept to clinical practice to clinical benefit. Am Heart J 2004;148:S19-S26.
- Suzuki H, Swei A, Zweifach BW, Schmid-Schonbein GW. In vivo evidence for microvascular oxidative stress in spontaneously hypertensive rats. Hydroethidine microfluorography. Hypertension 1995;25:1083-1089.
- [106] Swei A, Lacy F, Delano FA, Parks DA, Schmid-Schonbein GW. A mechanism of oxygen free radical production in the Dahl hypertensive rat. Microcirculation 1999;6:179-187.
- [107] Lenda DM, Sauls BA, Boegehold MA. Reactive oxygen species may contribute to reduced endothelium-dependent dilation in rats fed high salt. Am J Physiol Heart Circ Physiol 2000;279:H7-H14.
- [108] de Nigris F, Lerman LO, Condorelli M, Lerman A, Napoli C. Oxidation-sensitive transcription factors and molecular mechanisms in the arterial wall. Antioxid Redox Signal 2001;3:1119-1130.

- [109] Rathaus M, Bernheim J. Oxygen species in the microvascular environment: regulation of vascular tone and the development of hypertension. Nephrol Dial Transplant 2002;17:216-221.
- [110] Higashi Y, Sasaki S, Nakagawa K, Matsuura H, Oshima T, Chayama K. Endothelial function and oxidative stress in renovascular hypertension. N Engl J Med 2002;346: 1954-1962.
- [111] Frisbee JC, Roman RJ, Falck JR, Linderman JR, Lombard JH. Impairment of flow-induced dilation of skeletal muscle arterioles with elevated oxygen in normotensive and hypertensive rats. Microvasc Res 2000;60:37-48.
- [112] Yang D, Feletou M, Boulanger CM, Wu HF, Levens N, Zhang JN, Vanhoutte PM. Oxygen-derived free radicals mediate endothelium-dependent contractions to acetylcholine in aortas from spontaneously hypertensive rats. Br J Pharmacol 2002;136:104-110.
- [113] DeLano FA, Parks DA, Ruedi JM, Babior BM, Schmid-Schonbein GW. Microvascular display of xanthine oxidase and NADPH oxidase in the spontaneously hypertensive rat. Microcirculation 2006;13:551-566.
- [114] Wallwork CJ, Parks DA, Schmid-Schonbein GW. Xanthine oxidase activity in the dexamethasone-induced hypertensive rat. Microvasc Res 2003;66:30-37.
- [115] Zhu XY, Daghini E, Chade AR, Rodriguez-Porcel M, Napoli C, Lerman A, Lerman LO. Role of oxidative stress in remodeling of the myocardial microcirculation in hypertension. Arterioscler Thromb Vasc Biol 2006;26: 1746-1752.
- [116] Rodriguez-Porcel M, Zhu XY, Chade AR, Amores-Arriaga B, Caplice NM, Ritman EL, Lerman A, Lerman LO. Functional and structural remodeling of the myocardial microvasculature in early experimental hypertension. Am J Physiol Heart Circ Physiol 2006;290:H978-H984.
- [117] Rodriguez-Porcel M, Lerman A, Herrmann J, Schwartz RS, Sawamura T, Condorelli M, Napoli C, Lerman LO. Hypertension exacerbates the effect of hypercholesterolemia on the myocardial microvasculature. Cardiovasc Res 2003;58:213-221.
- [118] Rodriguez-Porcel M, Lerman LO, Herrmann J, Sawamura T, Napoli C, Lerman A. Hypercholesterolemia and hypertension have synergistic deleterious effects on coronary endothelial function. Arterioscler Thromb Vasc Biol 2003:23:885-891.
- [119] Rodriguez-Porcel M, Herrman J, Chade AR, Krier JD, Breen JF, Lerman A, Lerman LO. Long-term antioxidant intervention improves myocardial microvascular function in experimental hypertension. Hypertension 2004;43:493-498.
- [120] Zhu XY, Daghini E, Chade AR, Napoli C, Ritman EL, Lerman A, Lerman LO. Simvastatin prevents coronary microvascular remodeling in renovascular hypertensive pigs. J Am Soc Nephrol 2007;18:1209-1217.
- [121] Booth GL, Kapral MK, Fung K, Tu JV. Relation between age and cardiovascular disease in men and women with diabetes compared with non-diabetic people: a populationbased retrospective cohort study. Lancet 2006;368:29-36.
- [122] Mäkimattila S, Virkamäki A, Groop PH, Cockcroft J, Utriainen T, Fagerudd J, Yki-Järvinen H. Chronic hyperglycemia impairs endothelial function and insulin sensitivity via different mechanisms in insulin-dependent diabetes mellitus. Circulation 1996;94:1276-1282.
- [123] Balletshofer BM, Rittig K, Enderle MD, Volk A, Maerker E, Jacob S, Matthaei S, Rett K, Häring HU. Endothelial dysfunction is detectable in young normotensive first-degree relatives of subjects with type 2 diabetes in association with insulin resistance. Circulation 2000;101:1780-1784.
- [124] Steinberg HO, Chaker H, Learning R, Johnson A, Brechtel G, Baron AD. Obesity/insulin resistance is associated with



- endothelial dysfunction. Implications for the syndrome of insulin resistance. J Clin Invest 1996;97:2601-2610.
- [125] De Vriese AS, Verbeuren TJ, Van de Voorde J, Lameire NH, Vanhoutte PM. Endothelial dysfunction in diabetes. Br J Pharmacol 2000;130:963-974.
- [126] Li H, Gutterman DD, Rusch NJ, Bubolz A, Liu Y. Nitration and functional loss of voltage-gated K+ channels in rat coronary microvessels exposed to high glucose. Diabetes 2004;53:2436-2442.
- [127] Miura H, Wachtel RE, Loberiza FR, Saito T, Miura M, Nicolosi AC, Gutterman DD. Diabetes mellitus impairs vasodilation to hypoxia in human coronary arterioles: reduced activity of ATP-sensitive potassium channels. Circ Res 2003;92:151-158.
- [128] Zeng G, Nystrom FH, Ravichandran LV, Cong LN, Kirby M, Mostowski H, Quon MJ. Roles for insulin receptor, PI3kinase, and Akt in insulin-signaling pathways related to production of nitric oxide in human vascular endothelial cells. Circulation 2000;101:1539-1545.
- [129] He Z, Opland DM, Way KJ, Ueki K, Bodyak N, Kang PM, Izumo S, Kulkarni RN, Wang B, Liao R, Kahn CR, King GL. Regulation of vascular endothelial growth factor expression and vascularization in the myocardium by insulin receptor and PI3K/Akt pathways in insulin resistance and ischemia. Arterioscler Thromb Vasc Biol 2006;26:787-793.
- [130] Nicolls MR, Haskins K, Flores SC. Oxidant stress, immune dysregulation, and vascular function in type I diabetes. Antioxid Redox Signal 2007;9:879-889.
- [131] Shah S, Iqbal M, Karam J, Salifu M, McFarlane SI. Oxidative stress, glucose metabolism, and the prevention of type 2 diabetes: pathophysiological insights. Antioxid Redox Signal 2007;9:911-929.
- [132] Liguori A, Abete P, Hayden JM, Cacciatore F, Rengo F, Ambrosio G, Bonaduce D, Condorelli M, Reaven PD, Napoli C. Effect of glycaemic control and age on lowdensity lipoprotein susceptibility to oxidation in diabetes mellitus type 1. Eur Heart J 2001;22:2075–2084.
- [133] Pieper GM, Siebeneich W. Oral administration of the antioxidant, N-acetylcysteine, abrogates diabetes-induced endothelial dysfunction. J Cardiovasc 1998;32:101-105.
- [134] Skyrme-Jones RA, O'Brien RC, Berry KL, Meredith IT. Vitamin E supplementation improves endothelial function in type I diabetes mellitus: a randomized, placebo-controlled study. J Am Coll Cardiol 2000;36:94-102.
- [135] Ting HH, Timimi FK, Boles KS, Creager SJ, Ganz P, Creager MA. Vitamin C improves endothelium-dependent vasodilation in patients with non-insulin-dependent diabetes mellitus. J Clin Invest 1996;97:22-28.
- [136] de Nigris F, Balestrieri ML, Williams-Ignarro S, D'Armiento FP, Fiorito C, Ignarro LJ, Napoli C. The influence of pomegranate fruit extract in comparison to regular pomegranate juice and seed oil on nitric oxide and arterial function in obese Zucker rats. Nitric Oxide 2007;17:50-54.
- [137] Griendling KK, FitzGerald GA. Oxidative stress and cardiovascular injury: part II: animal and human studies. Circulation 2003;108:2034-2040.
- [138] Mullen MJ, Wright D, Donald AE, Thorne S, Thomson H, Deanfield IE. Atorvastatin but not L-arginine improves endothelial function in type I diabetes mellitus: a doubleblind study. J Am Coll Cardiol 2000;36:410-416.
- [139] Tsunekawa T, Hayashi T, Kano H, Sumi D, Matsui-Hirai H, Thakur NK, Egashira K, Iguchi A. Cerivastatin, a hydroxymethylglutaryl coenzyme a reductase inhibitor, improves endothelial function in elderly diabetic patients within 3 days. Circulation 2001;104:376-379.
- [140] Eltzschig HK, Collard CD. Vascular ischaemia and reperfusion injury. Br Med Bull 2004;70:71-86.

- [141] Meredith IT, Currie KE, Anderson TJ, Roddy MA, Ganz P, Creager MA. Postischemic vasodilation in human forearm is dependent on endothelium-derived nitric oxide. Am J Physiol 1996;270:1435-14440.
- [142] Sternbergh WC, Makhoul RG, Adelman B. Nitric oxidemediated, endothelium-dependent vasodilation is selectively attenuated in the postischemic extremity. Surgery 1993;114:960-967.
- [143] Davenpeck KL, Guo JP, Lefer AM. Pulmonary artery endothelial dysfunction following ischemia and reperfusion of the rabbit lung. J Vasc Res 1993;30:145-153.
- [144] Stauton M, Drexler C, Dulitz MG, Ekbom DC, Schmeling WT, Farber NE. Effects of hypoxia-reoxygenation on microvascular endothelial function in the rat hippocampal slice. Anesthesiology 1999;91:1462-1469.
- [145] Hein TW, Zhang C, Wang W, Chang CI, Thengchaisri N, Kuo L. Ischemia-reperfusion selectively impairs nitric oxidemediated dilation in coronary arterioles: counteracting role of arginase. FASEB J 2003;17:2328-23230.
- [146] DeFily DV. Control of microvascular resistance in physiological conditions and reperfusion. J Mol Cell Cardiol 1998;30:2547-2554.
- [147] Stewart DJ, Pohl U, Bassenge E. Free radicals inhibit endothelium-dependent dilation in the coronary resistance bed. Am J Physiol 1988;255:H765-H769.
- [148] Tiefenbacher CP, Chilian WM, Mitchell M, DeFily DV. Restoration of endothelium-dependent vasodilation after reperfusion injury by tetrahydrobiopterin. Circulation 1996;94:1423-1429.
- [149] Banda MA, Lefer DJ, Granger DN. Postischemic endothelium-dependent vascular reactivity is preserved in adhesion molecule-deficient mice. Am J Physiol 1997;273: 2721-2725.
- [150] Harris NR, Granger DN. Neutrophil enhancement of reperfusion-induced capillary fluid filtration associated with hypercholesterolemia. Am J Physiol 1996;271: 1755-1761.
- [151] Harris NR. Opposing effects of L-NAME on capillary filtration rate in the presence or absence of neutrophils. Am J Physiol 1997;273:1320-1325.
- [152] Reffelmann T, Hale SL, Dow JS, Kloner RA. No-reflow phenomenon persists long-term after ischemia/reperfusion in the rat and predicts infarct expansion. Circulation 2003;108:2911-2917.
- [153] Zimmerman GA, McIntyre TM, Prescott SM, Stafforini DM. The platelet-activating factor signaling system and its regulators in syndromes of inflammation and thrombosis. Crit Care Med 2002;30:S294-S301.
- [154] Kurose I, Argenbright LW, Wolf R, Lianxi L, Granger DN. Ischemia/reperfusion-induced microvascular dysfunction: role of oxidants and lipid mediators. Am J Physiol 1997;272:2976-2982.
- [155] Lehr HA, Bittinger F, Kirkpatrick CJ. Microcirculatory dysfunction in sepsis: a pathogenetic basis for therapy? J Pathol 2000;190:373-386.
- [156] Lam C, Tyml K, Martin C, Sibbald W. Microvascular perfusion is impaired in a rat model of normotensive sepsis. I Clin Invest 1994;94:2077-2083.
- [157] Piper RD, Pitt-Hyde M, Li F, Sibbald WJ, Potter RF. Microcirculatory changes in rat skeletal muscle in sepsis. Am J Respir Crit Care Med 1996;154:931-937.
- [158] Drazenovic R, Samsel RW, Wylam ME, Doerschuk CM, Schumacker PT. Regulation of perfused capillary density in canine intestinal mucosa during endotoxemia. J Appl Physiol 1992;72:259-265.
- [159] Farquhar I, Martin CM, Lam C, Potter R, Ellis CG, Sibbald WJ. Decreased capillary density in vivo in bowel mucosa of rats with normotensive sepsis. J Surg Res 1996;61:190-196.



- [160] De Backer D, Creteur J, Preiser JC, Dubois MJ, Vincent JL. Microvascular blood flow is altered in patients with sepsis. Am J Respir Crit Care Med 2002;166:98-104.
- [161] Ellis CG, Jagger J, Sharpe M. The microcirculation as a functional system. Crit Care 2005;9(Suppl 4):S3-S8.
- [162] Cerwinka WH, Cooper D, Krieglstein CF, Ross CR, McCord JM, Granger DN. Superoxide mediates endotoxin-induced platelet-endothelial cell adhesion in intestinal venules. Am J Physiol Heart Circ Physiol 2003;284: H535-H541.
- [163] Martins PS, Kallas EG, Neto MC, Dalboni MA, Blecher S, Salomao R. Upregulation of reactive oxygen species generation and phagocytosis, and increased apoptosis in human neutrophils during severe sepsis and septic shock. Shock 2003;20:208-212.
- [164] Bateman RM, Sharpe MD, Ellis CG. Bench-to-bedside review: microvascular dysfunction in sepsis--hemodynamics, oxygen transport, and nitric oxide. Crit Care 2003;7: 359-373.
- Hollenberg SM, Easington CR, Osman J, Broussard M, Parrillo JE. Effects of nitric oxide synthase inhibition on microvascular reactivity in septic mice. Shock 1999;12: 262-267.
- [166] Kinugawa K, Takahashi T, Kohmoto O, Yao A, Aoyagi T, Momomura S, Hirata Y, Serizawa T. Nitric oxide-mediated effects of interleuking-6 on [Ca 2+] and cell contraction in cultured chick ventricular myocytes. Circ Res 1994;75: 285-295.
- [167] Kumar A, Brar R, Wang P, Dee L, Skorupa G, Khadour F, Schulz R, Parrillo JE. Role of nitric oxide and cGMP in

- human septic serum-induced depression of cardiac myocyte contractility. Am J Physiol 1999;276:R265-R276.
- [168] Stein B, Frank P, Schmitz W, Scholz H, Thoenes M. Endotoxin and cytokines induce direct cardiodepressive effects in mammalian cardiomyocytes via induction of nitric oxide synthase. J Mol Cell Cardiol 1996;28:1631-1639.
- [169] Ullrich R, Scherrer-Crosbie M, Bloch KD, Ichinose F, Nakajima H, Picard MH, Zapol WM, Quezado ZM. Congenital deficiency of nitric oxide synthase 2 protects against endotoxin-induced myocardial dysfunction in mice. Circulation 2000;102:1440-1446.
- [170] Strunk V, Hahnenkamp K, Schneuing M, Fischer LG, Rich GF. Selective iNOS inhibition prevents hypothension in septic rats while preserving endothelium-dependent vasodilation. Anesth Analg 2001;92:681-682.
- [171] Revelly JP, Ayuse T, Brienza N, Fessler HE, Robotham JL. Endotoxic shock alters distribution of blood flow within the intestinal wall. Crit Care Med 1996;24:1345-1351.
- [172] Crimi E, Sica V, Slutsky AS, Zhang H, Williams-Ignarro S, Ignarro LJ, Napoli C. Role of oxidative stress in experimental sepsis and multisystem organ dysfunction. Free Radic Res 2006:40:665-672.
- [173] Ichinose F, Buys ES, Neilan TG, Furutani EM, Morgan JG, Jassal DS, Graveline AR, Searles RJ, Lim CC, Kaneki M, Picard MH, Scherrer-Crosbie M, Janssens S, Liao R, Bloch KD. Cardiomyocyte-specific overexpression of nitric oxide synthase 3 prevents myocardial dysfunction in murine models of septic shock. Circ Res 2007;100:130-139.
- [174] Napoli C, Maione C, Schiano C, Fiorito C, Ignarro LJ. Bone marrow cell-mediated cardiovascular repair: potential of combined therapies. Trends Mol Med 2007;13:278-286.

